



# OAT

## ORGANIC ACIDS TEST

### Summary of Results

REQUISITION #	9900002	COLLECTION TIME	06:30 AM
PATIENT NAME	Report Sample	COLLECTION DATE	Apr 5, 2025
DATE OF BIRTH	Mar 9, 1960	SAMPLE TYPE	Urine
GENDER F	AGE 65	REPORT DATE	Mar 4, 2026
PRACTITIONER	NO PHYSICIAN		

KEY

IMBALANCE



UNREMARKABLE



MARGINAL



SIGNIFICANT

#### Microbial Overgrowth

Results may indicate significant microbial imbalance.



- 7 Arabinose
- 6 Tartaric
- 5 Furancarboxylglycine
- 21 Oxalic

11A

#### Mitochondrial Health

Results may indicate significant imbalances.



- 24 Succinic
- 46 Methylsuccinic
- 48 Suberic
- 53 Glutaric

13B

#### Neurotransmitter Metabolites

Results may indicate marginal imbalances in neurotransmitter metabolites.



- 36 Dihydroxyphenylacetic (DOPAC)
- 54 Ascorbic
- 53 Glutaric
- 50 Methylmalonic

2C

#### Toxic Exposure

Results may indicate significant toxic exposure.



- |                        |                 |
|------------------------|-----------------|
| 5 Furancarboxylglycine | 58 Pyroglutamic |
| 6 Tartaric             | 21 Oxalic       |
| 9 Tricarballic         | 29 Citric       |
| 24 Succinic            |                 |

9D

#### Methylation/Detoxification

Results may indicate significant detoxification and/or methylation imbalances.



- 58 Pyroglutamic
- 50 Methylmalonic
- 53 Glutaric

11E

#### Nutrient Needs

Results may indicate multiple nutrient needs.



- |                   |                                  |
|-------------------|----------------------------------|
| 53 Glutaric       | 36 Dihydroxyphenylacetic (DOPAC) |
| 21 Oxalic         | 9 Tricarballic                   |
| 46 Methylsuccinic | 58 Pyroglutamic                  |
| 48 Suberic        |                                  |

16F



METABOLITE

REFERENCE RANGE (mmol/mol creatinine)

RESULTS

REFERENCE POPULATION  
Females Age 13 and Over

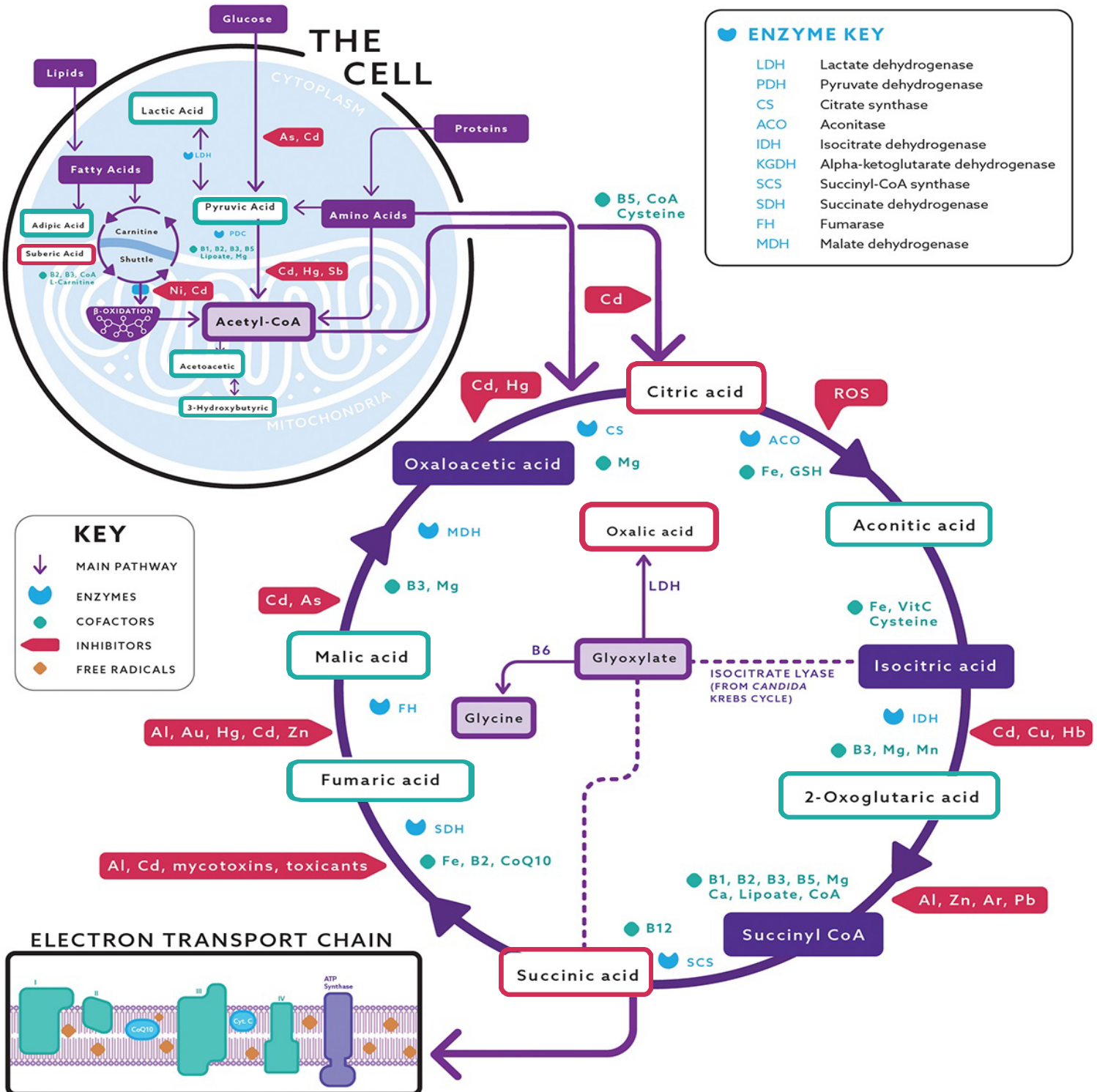
INTESTINAL MICROBIAL OVERGROWTH

YEAST AND FUNGAL

BACTERIAL

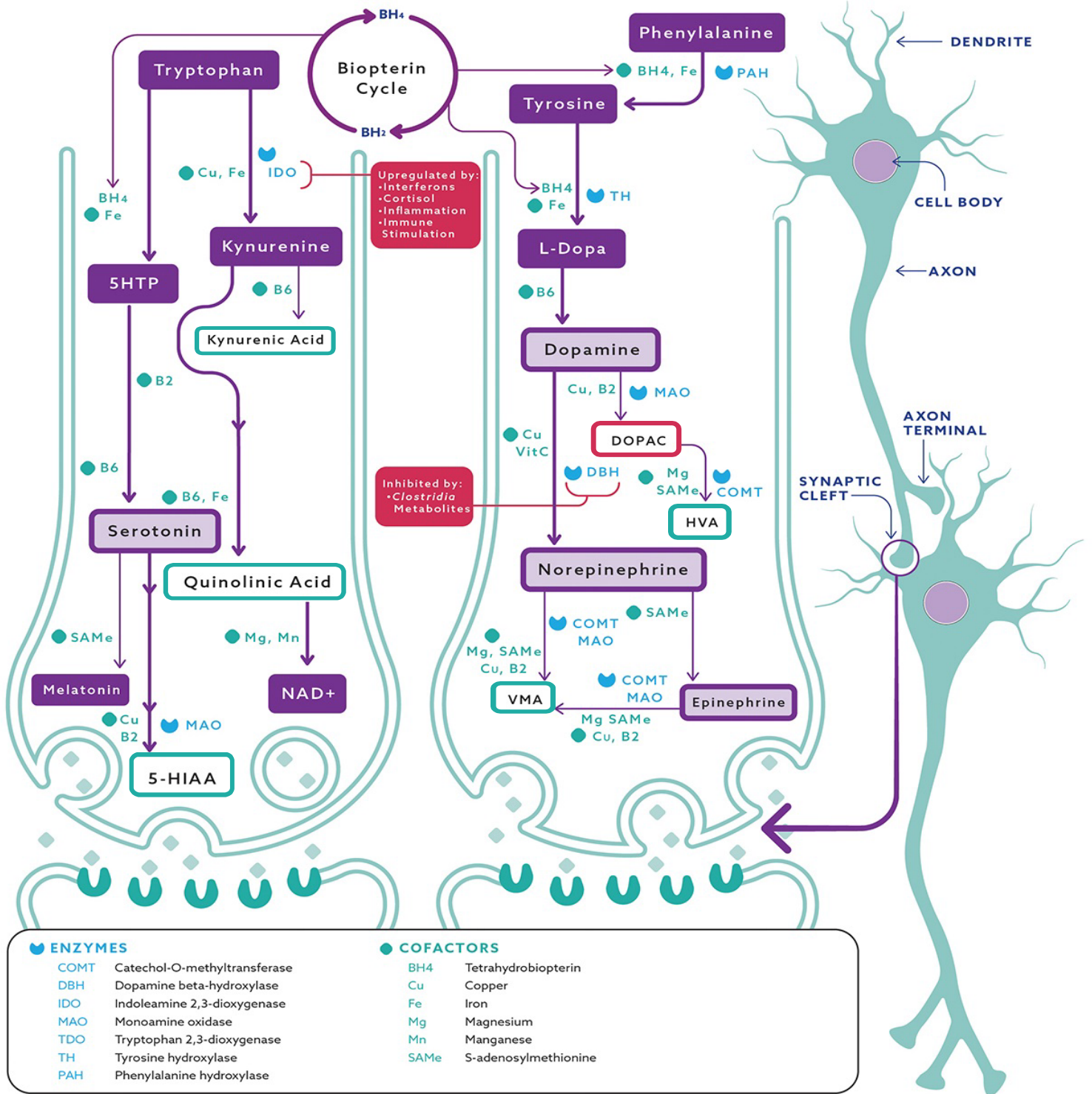
CLOSTRIDIA BACTERIAL

Metabolite	Reference Range	Result	Reference Population
1 Citramalic	≤3.6	1.6	1.6
2 5-Hydroxymethyl-2-furoic (Aspergillus)	≤14	14	14
3 3-Oxoglutaric	≤0.33	0.23	0.23
4 Furan-2,5-dicarboxylic (Aspergillus)	≤16	8.1	8.1
5 Furancarboxylglycine (Aspergillus)	≤1.9	H 15	15
6 Tartaric (Aspergillus)	≤4.5	H 6.2	6.2
7 Arabinose	≤29	H 69	69
8 Carboxycitric	≤29	12	12
9 Tricarballic (Fusarium)	≤0.44	H 0.55	0.55
10 Hippuric	≤613	H 1,340	1340
11 2-Hydroxyphenylacetic	0.06 - 0.66	0.53	0.53
12 4-Hydroxybenzoic	≤1.3	1.2	1.2
13 4-Hydroxyhippuric	0.79 - 17	8.7	8.7
14 DHPPA (Beneficial Bacteria)	≤0.38	H 0.57	0.57
15 4-Hydroxyphenylacetic (C. difficile, C. stricklandii & others)	≤19	15	15
16 HPPHA (C. sporogenes, C. botulinum & others)	≤208	162	162
17 4-Cresol (C. difficile)	≤75	37	37
18 3-Indoleacetic (C. stricklandii, C. subterminale & others)	≤11	2.9	2.9





METABOLITE	REFERENCE RANGE	RESULTS (mmol/mol creatinine)	REFERENCE POPULATION (Females Age 13 and Over)	
<b>OXALATE METABOLITES</b>				
19 Glyceric	0.77 - 7.0	H 7.6		
20 Glycolic	16 - 117	89		
21 Oxalic	6.8 - 101	H 224		
<b>GLYCOLYTIC METABOLITES</b>				
22 Lactic	≤48	22		
23 Pyruvic	≤9.1	2.4		
<b>MITOCHONDRIAL MARKERS</b>				
CITRIC ACID CYCLE	24 Succinic	≤9.3	H 18	
	25 Fumaric	≤0.94	0.82	
	26 Malic	0.06 - 1.8	1.7	
	27 2-Oxoglutaric	≤35	11	
	28 Aconitic	6.8 - 28	14	
	29 Citric	≤507	H 610	
<b>MITOCHONDRIAL MARKERS</b>				
AMINO ACIDS	30 3-Methylglutaric	≤0.76	0.35	
	31 3-Hydroxyglutaric	≤6.2	5.4	
	32 3-Methylglutaconic	≤4.5	1.4	
<b>NEUROTRANSMITTER METABOLITES</b>				
PHENYLALANINE AND TYROSINE	33 Homovanillic (HVA) (dopamine)	0.80 - 3.6	3.5	
	34 Vanillylmandelic (VMA) (norepinephrine, epinephrine)	0.46 - 3.7	2.5	
	35 HVA / VMA Ratio	0.16 - 1.8	1.4	
	36 Dihydroxyphenylacetic (DOPAC) (dopamine)	0.08 - 3.5	H 4.6	
	37 HVA/ DOPAC Ratio	0.10 - 1.8	0.77	
TRYPTOPHAN	38 5-Hydroxyindoleacetic (5-HIAA) (serotonin)	≤4.3	1.9	
	39 Quinolinic	0.85 - 3.9	2.4	
	40 Kynurenic	≤2.2	1.4	





METABOLITE

REFERENCE RANGE

RESULTS (mmol/mol creatinine)

REFERENCE POPULATION  
Females Age 13 and Over

PYRIMIDINE METABOLITES - FOLATE METABOLISM

④1	Uracil	≤9.7	4.5	
④2	Thymine	≤0.56	0.19	

KETONE AND FATTY ACID OXIDATION

④3	3-Hydroxybutyric	≤3.1	1.7	
④4	Acetoacetic	≤10	1.3	
④5	Ethylmalonic	0.44 - 2.8	2.1	
④6	Methylsuccinic	0.10 - 2.2	<b>H</b> 4.1	
④7	Adipic	0.04 - 3.8	2.0	
④8	Suberic	0.18 - 2.2	<b>H</b> 3.2	
④9	Sebacic	≤0.24	0.21	

NUTRITIONAL MARKERS

Vitamin B12

⑤0	Methylmalonic *	≤2.3	<b>H</b> 2.8	
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Vitamin B6

⑤1	Pyridoxic	≤34	3.7	
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Vitamin B5

⑤2	Pantothenic	≤10	<b>H</b> 23	
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Vitamin B2 (Riboflavin)

⑤3	Glutaric *	0.04 - 0.36	<b>H</b> 0.89	
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Vitamin C

⑤4	Ascorbic	10 - 200	<b>L</b> 0.56	
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Vitamin Q10 (CoQ10)

⑤5	3-Hydroxy-3-methylglutaric *	0.17 - 39	29	
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Glutathione Precursor and Chelating Agent

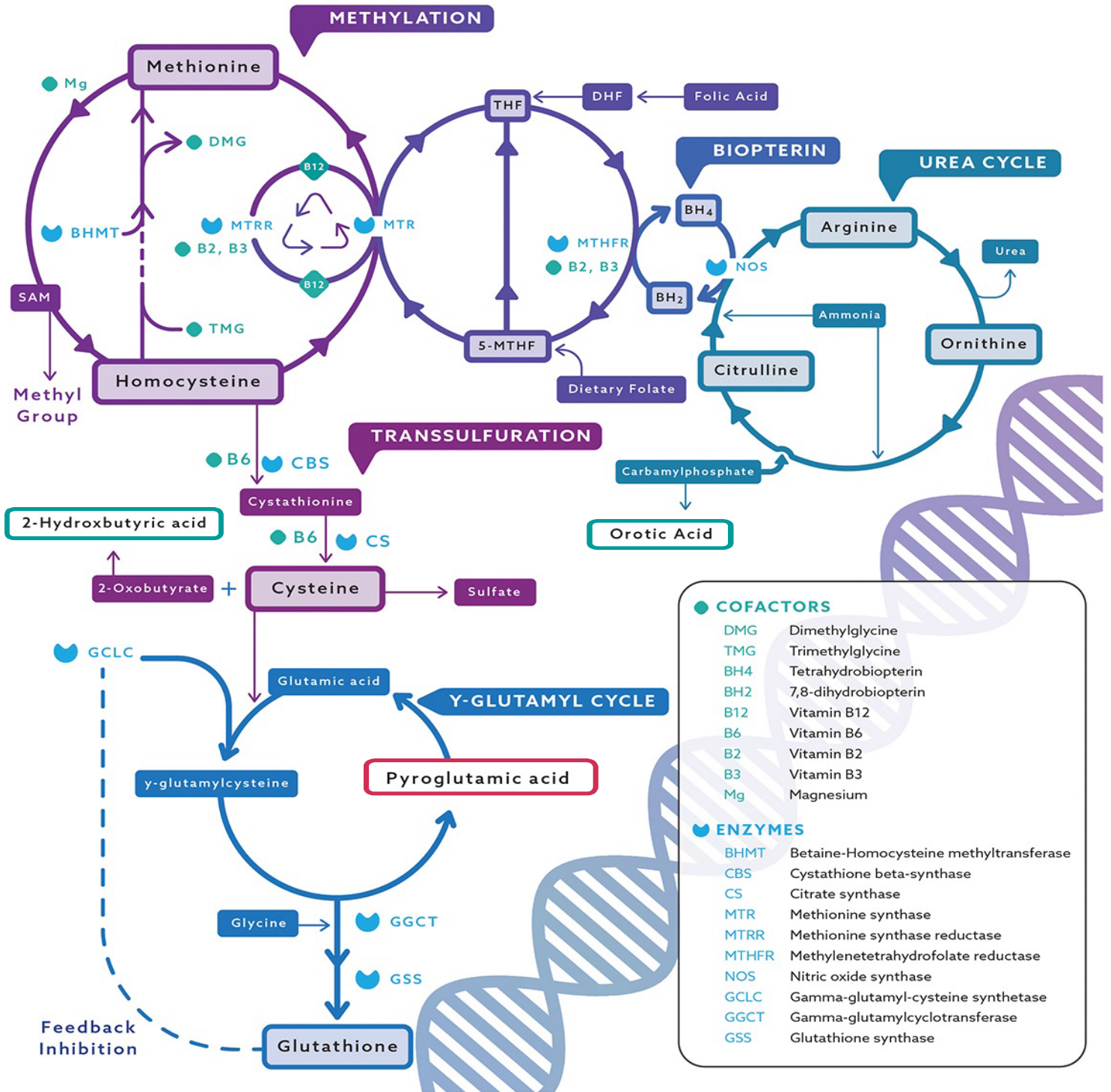
⑤6	N-Acetylcysteine (NAC)	≤0.28	0.04	
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Biotin (Vitamin H)

⑤7	Methylcitric *	0.19 - 2.7	1.1	
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\* A high value for this marker may indicate a deficiency of this vitamin.







METABOLITE

REFERENCE RANGE

RESULTS (mmol/mol creatinine)

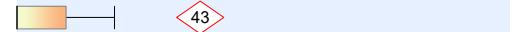
REFERENCE POPULATION  
Females Age 13 and Over

INDICATORS OF DETOXIFICATION

Glutathione

58 Pyroglutamic \*

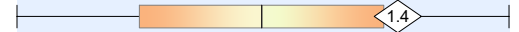
10 - 33 H 43



Methylation, Toxic exposure

59 2-Hydroxybutyric \*\*

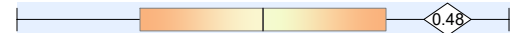
0.03 - 1.8 1.4



Ammonia Excess

60 Orotic

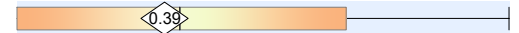
0.06 - 0.54 0.48



Aspartame, Salicylates, or GI Bacteria

61 2-Hydroxyhippuric

≤ 1.3 0.39



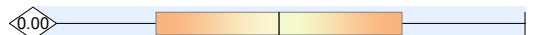
\* A high value for this marker may indicate a Glutathione deficiency. \*\* High values may indicate methylation defects and/or toxic exposure.

AMINO ACID METABOLITES

62 2-Hydroxyisovaleric

≤ 2.0

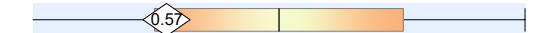
0



63 2-Oxoisovaleric

≤ 2.1

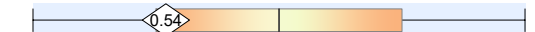
0.57



64 3-Methyl-2-oxovaleric

≤ 2.0

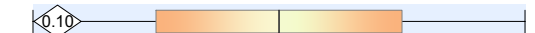
0.54



65 2-Hydroxyisocaproic

≤ 2.0

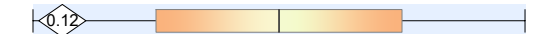
0.10



66 2-Oxoisocaproic

≤ 2.0

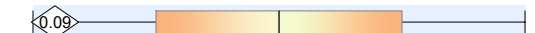
0.12



67 2-Oxo-4-methiolbutyric

≤ 2.0

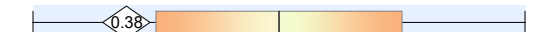
0.09



68 Mandelic

≤ 2.0

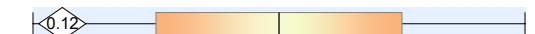
0.38



69 Phenyllactic

≤ 2.0

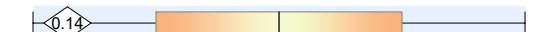
0.12



70 Phenylpyruvic

≤ 2.0

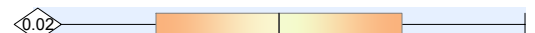
0.14



71 Homogentisic

≤ 2.0

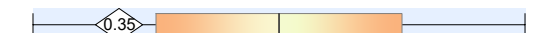
0.02



72 4-Hydroxyphenyllactic

≤ 2.0

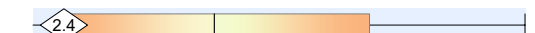
0.35



73 N-Acetylaspartic

≤ 38

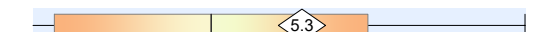
2.4



74 Malonic

≤ 9.7

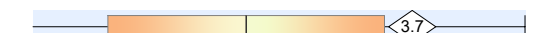
5.3



75 4-Hydroxybutyric

≤ 4.8

3.7



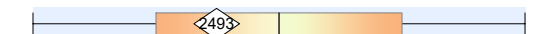
Low values are not necessarily associated with inadequate protein intake and have not been demonstrated to indicate specific amino acid deficiencies.

MINERAL METABOLISM

76 Phosphoric

1,000 - 5,000

2,493



INDICATOR OF FLUID INTAKE

77 Creatinine \*

10

mg/dL

\* The sample's creatinine level is below 20 mg/dL, which may affect the interpretation of creatinine-normalized results; interpret results with caution.



The reference ranges for organic acids were established using samples collected from typical individuals of all ages with no known physiological or psychological disorders. The ranges were determined by calculating the mean and standard deviation (SD) and are defined as + 2SD of the mean. Reference ranges are age and gender specific, consisting of Male Adult ( $\geq 13$  years), Female Adult ( $\geq 13$  years), Male Child ( $< 13$  years), and Female Child ( $< 13$  years).

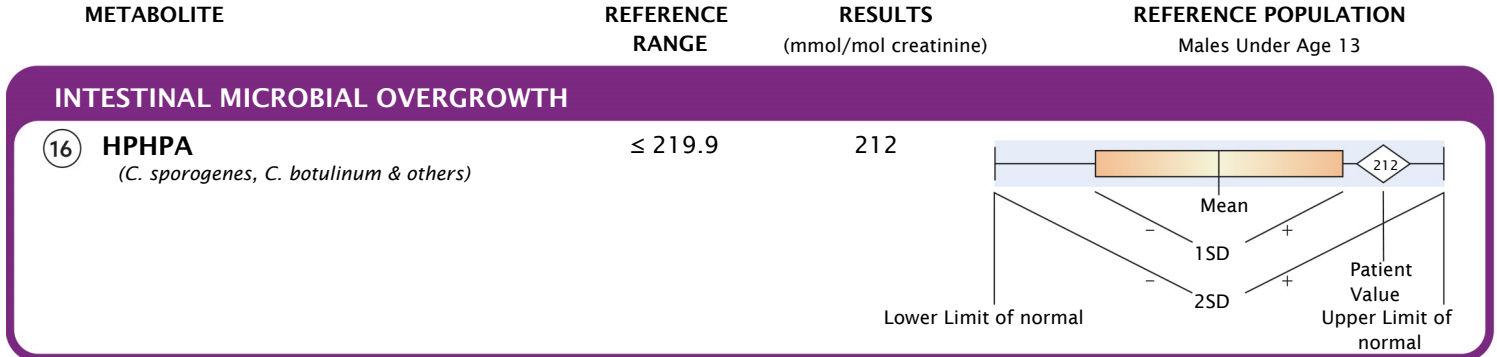
There are two types of graphical representations of patient values in the report format of both the standard Organic Acids Test and the Microbial Organic Acids Test.

The first graph will occur when the value of the patient is within the reference (normal) range, defined as the mean plus or minus two standard deviations.

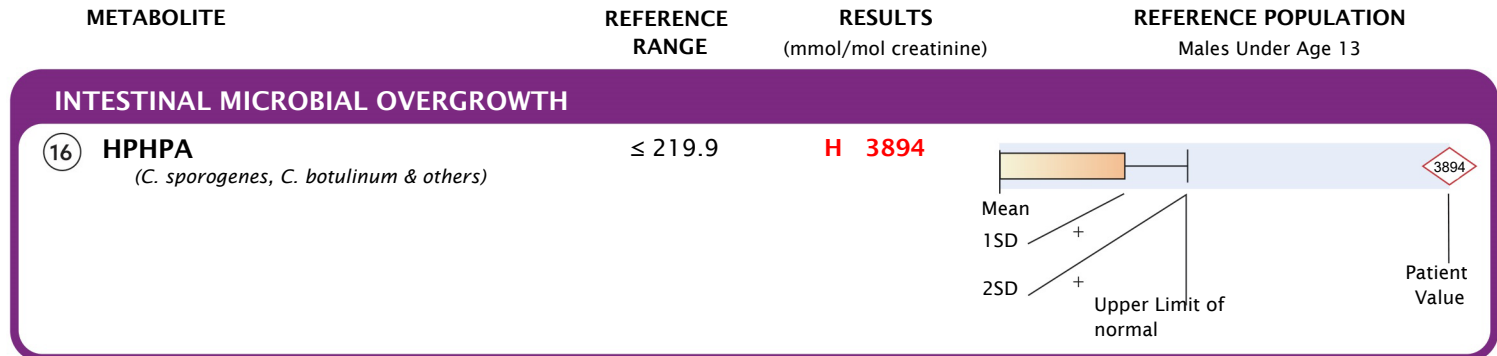
The second graph will occur when the value of the patient exceeds the upper limit of normal. In such cases, the graphical reference range is "shrunk" so that the degree of abnormality can be appreciated at a glance. In this case, the lower limits of normal are not shown, only the upper limit of normal is shown.

In both cases, the value of the patient is given to the left of the graph and is repeated on the graph inside a diamond. If the value is within the normal range, the diamond will be outlined in black. If the value is high or low, the diamond will be outlined in red.

### EXAMPLE OF VALUE WITHIN REFERENCE RANGE



### EXAMPLE OF ELEVATED VALUE



Methodology: GC-MS.Creatinine by Jaffe Reaction. The creatinine test is performed to adjust metabolic marker results for differences in fluid intake. Urinary creatinine, from a random collection, has limited diagnostic value due to variability as a result of recent fluid intake.



For more extensive interpretations, refer to the [support guide](#)

ICON KEY



Microbial Overgrowth



Neurotransmitter Metabolites



Methylation/Detoxification



Mitochondrial Health



Toxic Exposure



Nutrient Needs

## 5 Furancarboxylglycine



Furancarboxylglycine, also known as 2-furoylglycine, is a furan-derived acyl glycine that may be linked to the fungus *Aspergillus*, serving as a marker for mold exposure or overgrowth. Additionally, dietary intake of high-temperature processed foods and beverages, such as coffee, can influence its levels.

## 6 Tartaric



Tartaric acid is a naturally occurring dicarboxylic acid associated with the activity of *Aspergillus*, *Penicillium*, and to a lesser extent, *Candida* and *Saccharomyces*. Elevated levels may indicate fungal dysbiosis. It can also inhibit the Krebs cycle by disrupting malic acid utilization, potentially impacting mitochondrial function. Additionally, dietary sources such as grapes, red wine, tamarind, and certain food additives may contribute to elevated levels.

## 7 Arabinose



Arabinose is an aldopentose sugar that can be metabolized by various organisms through pathways involving conversion into intermediates of the pentose phosphate pathway. It has been studied as a marker for intestinal yeast overgrowth and shown clinically to respond to antifungal therapies. Arabinose is also a naturally occurring sugar compound found in numerous plants, such as beets and grains.



9 Tricarballic

Tricarballic acid is often associated with the fumonisin class of mycotoxins produced mainly by various *Fusarium* species and *Aspergillus nigri* (black aspergilli). These mold exposures can come from indoor environments and contaminated dietary sources such as numerous grain products. This metabolite may also be produced from certain bacteria. Tricarballic acid has been shown to bind to various minerals, influencing nutritional needs.



10 Hippuric

Hippuric acid is a conjugate of glycine and benzoic acid, and may be influenced by gut bacteria, diet, and environmental exposures. It can be produced during the breakdown of benzene-type aromatic compounds from either microbial activity in the gut, exposure to toluene, or naturally produced in the liver from dietary intake of polyphenol-rich foods. Low levels can indicate poor microbial activity, glycine or B5 insufficiencies, and has also been associated with numerous chronic conditions. In rare cases, extreme elevations may be linked to genetic metabolic disorders.



14 DHPPA

3,4-Dihydroxyphenylpropionic acid (DHPPA), also known as dihydrocaffeic acid, is a metabolite produced by gut microbiota through the breakdown of certain dietary polyphenols. DHPPA is primarily associated with *Lactobacilli*, *Bifidobacteria*, *E. coli*, and some *Clostridium* species often identified as commensal. Elevated levels may indicate a polyphenol-rich diet or an abundance of these flora, while low levels suggest insufficient polyphenols or potentially reduced beneficial bacteria.



19 Glyceric

Elevated glyceric acid can result from microbial imbalances, nutrient deficiencies, diet, or genetic mutations. Elevated glyceric acid from fungal or microbial overgrowth, may be through the glyoxylate pathway, where excess glyoxylate is converted to glyceric acid or from direct production. Insufficiencies in vitamin B3 or tryptophan, which are cofactors for glyoxylate reductase/hydroxypyruvate reductase (GRHPR), the enzyme regulating this pathway, may also contribute to increased levels. Additionally, high fructose intake, glycerol metabolism, and various genetic conditions can also lead to excess glyceric acid. Elevated levels have also been associated with conditions such as glucose intolerance, rheumatoid arthritis, and psychiatric illnesses.



21 Oxalic

Oxalates, the anionic form of oxalic acid, are naturally occurring compounds that can form crystalline structures when combined with a cation (e.g. calcium, lead, etc). Various organisms such as plants and fungi produce them for structure, defense, and function, but they are also a normal part of human metabolism. Molds and yeasts can produce oxalates directly and indirectly, respectively, while beneficial bacteria such as *Oxalobacter formigenes* help break them down. Additionally, exposure to certain toxicants such as ethylene oxide, ethylene glycol, and trichloroacetic acid can increase levels in the urine. When consumed from oxalate-containing plants (e.g. spinach, soy, beets, tea. etc.), soluble and unbound oxalates are either absorbed in the colon or bind to minerals like calcium and form insoluble compounds in the intestinal lumen, followed by excretion in feces. Oxalates' ability to bind essential minerals may contribute to deficiencies, while factors like vitamin B6 insufficiency, fat malabsorption, high-oxalate food consumption, collagen supplementation, and certain medications can further impact oxalate excretion. Regardless of source, in humans, excessive oxalates can potentially cause systemic problems and have been associated with numerous health conditions involving pain and inflammation.



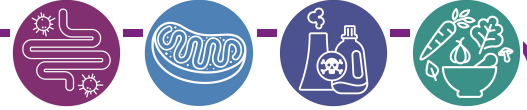
24 Succinic

Succinic acid plays a key role in the Krebs cycle and the electron transport chain, where it is oxidized to fumarate (fumaric acid) via succinate dehydrogenase (SDH), also known as mitochondrial Complex II. SDH is the only enzyme that participates in both the Citric Acid Cycle and the Electron Transport Chain (ETC), giving insights into both pathways. Elevated levels can indicate mitochondrial dysfunction, microbial imbalance, or toxic exposures, such as heavy metals or certain fungicides. Deficiencies in nutrients such as iron, riboflavin (B2), and CoQ10 can also contribute to increased succinic acid levels.





29 Citric



Citric acid is a key component of the Krebs cycle, formed from acetyl-CoA and oxaloacetate by the enzyme citrate synthase. The enzyme aconitase, which requires iron for activity, converts citric acid to isocitrate, and its function is essential for proper citric acid metabolism. Elevated citric acid levels can indicate microbial overgrowth, mitochondrial dysfunction, or toxic exposure to substances such as arsenic or aluminum. Nutrient deficiencies, particularly in iron, can reduce aconitase activity, leading to increased citric acid levels, while oxidative stress can also inhibit the enzyme. Low citric acid levels may result from conditions such as hypokalemia, impaired metabolism, or iron overload, and can increase the risk of oxalate stone formation due to greater free calcium availability.

36 Dihydroxyphenylacetic (DOPAC)



Dihydroxyphenylacetic acid (DOPAC) is a key dopamine metabolite formed through the oxidative deamination of dopamine by monoamine oxidase (MAO), resulting in a neurotoxic byproduct, 3,4-dihydroxyphenylacetaldehyde (DOPAL), which is further metabolized into DOPAC by aldehyde dehydrogenase enzyme (ALDH2). DOPAC, along with HVA and VMA, is used to assess dopamine and norepinephrine metabolism with fluctuations in these metabolites reflecting alterations in catecholamine turnover and balance. DOPAC levels can be elevated due to disruptions in dopamine metabolism, which can be caused by DBH inhibition via clostridia or toxic exposures such as mycotoxins or heavy metals. Nutritional deficiencies in vitamin B6, B2, magnesium, copper, and vitamin C, along with genetic variations in COMT or DBH, can also impact DOPAC levels. Low precursors like phenylalanine and tyrosine, as well as deficiencies in cofactors like tetrahydrobiopterin and other enzymatic cofactors, can impair dopamine synthesis and metabolism, leading to lower DOPAC levels. Additionally, dietary factors, medications, and certain pharmaceuticals can further affect dopamine metabolism and lead to either elevated or reduced DOPAC levels.

46 Methylsuccinic



Methylsuccinic acid (MSA), is a fatty acid metabolite involved in isoleucine metabolism. It is associated with mitochondrial dysfunction from dysfunction of fatty acid utilization, and has been associated with type 2 diabetes, and various inborn errors of metabolism.



48 Suberic

Suberic acid is a dicarboxylic acid that serves as a key marker of fatty acid metabolism that undergoes omega-oxidation when beta oxidation is impaired. High suberic acid may indicate mitochondrial dysfunction, carnitine or Vitamin B2 (Riboflavin) insufficiencies, metabolic stressors, dietary intake, or in rare cases, genetic abnormalities.

50 Methylmalonic



Vitamin B12 is essential for propionic acid metabolism, as it enables the conversion of methylmalonyl-CoA to succinyl-CoA through the B12-dependent enzyme methylmalonyl-CoA mutase (MMUT). When B12 is deficient, this process is disrupted, causing the accumulation of methylmalonic acid (MMA). Propionic acid metabolism connects branched-chain amino acid and odd-chain fatty acid oxidation to the Citric Acid Cycle, and disturbances in this pathway can have widespread metabolic consequences. Vitamin B12 not only plays a role in mitochondrial function, but it can also disrupt the methylation processes, which can have cascading effects, particularly in neurotransmitter metabolism, and detoxification pathways.

52 Pantothenic



Pantothenic acid (B5) plays a crucial role in metabolism, primarily through its involvement in Coenzyme A (CoA) synthesis, which is essential for energy production. Levels of B5 can be influenced by dietary intake, microbial activity, supplementation including products containing B5 derivatives or royal jelly. Rare genetic mutations, such as those in the PANK2 gene linked to PKAN, can lead to significantly elevated levels due to impaired conversion to CoA. Additionally, low B5 levels may have a significant influence on mitochondrial dysfunction, potentially affecting the Citric Acid Cycle and energy metabolism.

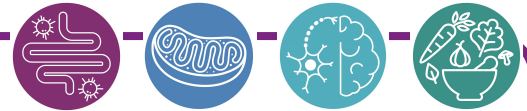


53 **Glutaric**



Glutaric acid metabolism relies on riboflavin (B2) as a cofactor for glutaryl-CoA dehydrogenase, and a deficiency can lead to its accumulation, impacting amino acid and fatty acid metabolism. Riboflavin is essential for mitochondrial energy production, neurotransmitter synthesis, and detoxification through its role in FAD-dependent enzymes. Insufficient B2 levels can affect ATP synthesis, methylation, and oxidative stress balance, potentially contributing to metabolic dysfunction. Dietary restrictions, medications such as oral contraceptives, and inadequate riboflavin intake may further influence these processes, increasing the risk of deficiency-related complications.

54 **Ascorbic**



Ascorbic acid, also known as vitamin C, is a water-soluble nutrient found in citrus fruits and vegetables, essential for collagen synthesis, various biological functions, and antioxidant protection. Ascorbic acid levels are influenced by factors such as microbial imbalance, mitochondrial health, neurotransmitter metabolism, and nutritional needs such as iron or zinc deficiency. Foods rich in vitamin C and supplements, as well as medications, can increase its levels, while conditions such as critical illness, infections, and smoking raise the body's demand for it. Vitamin C levels are commonly low, due to its ability to be rapidly metabolized and excreted as well as its short half-life.

58 **Pyroglutamic**



Pyroglutamic acid (5-oxoproline) is formed from glutamate, glutamine, and gamma-glutamylated peptides through gamma-glutamylcyclotransferase and is involved in glutathione metabolism. When intracellular glutathione synthesis is impaired, pyroglutamic acid levels rise, prompting increased cystine uptake to support restoration and reduce its accumulation. Factors such as microbial imbalance, mitochondrial dysfunction, toxic exposures, methylation, and nutritional deficiencies (e.g., magnesium, vitamin B6, precursor amino acids) can elevate pyroglutamic acid levels by disrupting glutathione metabolism and related cycles. Conversely, genetic mutations, severe nutritional deficiencies, and metabolic disorders may influence low levels, though this is mainly theoretical.



# OAT

ORGANIC ACIDS TEST

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